

**UNIVERSITY OF CAMBRIDGE INTERNATIONAL EXAMINATIONS**  
GCE Advanced Subsidiary Level and GCE Advanced Level

**MARK SCHEME for the October/November 2010 question paper  
for the guidance of teachers**

**9700 BIOLOGY**

**9700/51**

Paper 5 (Planning, Analysis and Evaluation),  
maximum raw mark 30

This mark scheme is published as an aid to teachers and candidates, to indicate the requirements of the examination. It shows the basis on which Examiners were instructed to award marks. It does not indicate the details of the discussions that took place at an Examiners' meeting before marking began, which would have considered the acceptability of alternative answers.

Mark schemes must be read in conjunction with the question papers and the report on the examination.

- CIE will not enter into discussions or correspondence in connection with these mark schemes.

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Mark schemes abbreviations:

**;** separates marking points

**/** alternative answers for the same point

**R** reject

**A** accept (for answers correctly cued by the question, or guidance on the mark scheme)

**AW** alternative wording (where responses vary more than usual)

**underline** actual word given must be used by the candidate (grammatical variants excepted)

**max** indicates the maximum number of marks that can be given

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| Question          | Expected answer   | Extra guidance   | Mark |
|-------------------|---|--|------|
| 1 (a) (i)<br>(ii) | <p>light + intensity / exposure;</p> <p>8 of:</p> <p><i>independent variable:</i></p> <p>1. ref. to a systematic way of obtaining leaves;</p> <p><i>dependent variables:</i></p> <p>2. ref. to a <b>method</b> of measuring surface area;</p> <p>3. ref. to how surface area is calculated;</p> <p>4. ref. to idea of both sides needed to get total surface area;</p> <p>5. ref. to a method of measuring mass;</p> <p>6. ref. to finding dry mass;</p> <p>7. ref. to a method of measuring internode length either on the plant or a cut section from a plant;</p> <p>8. ref. to a method of measuring water loss;</p> <p>9. ref. method of using the transpiration apparatus;</p> <p>10. ref. to keeping constant environment when measuring water loss;</p> <p>(max. 6)</p> <p><i>safety:</i></p> <p>11. ref. to low risk investigation;</p> <p><i>reliability</i></p> <p>12. ref. to mean values of the whole sample;</p> <p>13. ref. to method of working out SA : mass ratio;</p> <p>14. ref. to calculating standard deviation;</p> | <p>do not allow light unqualified or position in shade / sun</p><br><p><i>ignore any reference to planting seeds / potted plants</i></p> <p>1. e.g. 3<sup>rd</sup> leaf from the apex / different heights / all from the same height / equal light exposure</p> <p>2. e.g. draw round each leaf on grid or use transparent grid over leaf / measure <b>diameter(s)</b> of leaf</p> <p>3. count squares / use formula <math>\pi r^2</math></p> <p>5. e.g. digital balance / scales</p> <p>6. e.g. sample leaves dried in oven until mass constant</p> <p>7. by holding against a ruler / use string or cotton to mark distance measure with ruler</p> <p>8. e.g. use a potometer / weigh leaf / place leaf inside a plastic bag (to collect water)</p> <p>9. measure distance moved by water / weigh at hourly intervals / weigh bag or leaf after a stated time</p><br><p>11. e.g. ref. heat and suitable precaution if use dry mass / leaf allergy</p> <p>12. do not allow 'mean of three idea'</p> <p>14. ignore formula</p> | [1]  |
|                   |   |  | [8]  |

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| (b) (i) | $t = 23 - 15 ;$<br>$\sqrt{\frac{4^2}{30} + \frac{3^2}{30}} ;$<br>$\frac{(8)}{(0.9)} = 8.9;$   | <p>ignore any working in the answer</p> <p>allow 9 / 8.89 and 8.88 8</p> <p>allow ecf for incorrect figure from subtraction</p>                             | [3] |
| (ii)    | <p>total number of measurements –1 for each set of measurement /</p> $(30 - 1) + (30 - 1) = 58;$  | <p>allow</p> $2n - 2 / (n - 1) + (n - 1)$<br>$60 - 2 = 58$  | [1] |
| (iii)   | <p>ref. (both) calculated / <math>t</math> values are greater than the critical value / 0.2;</p> <p>both results are significant / not due to chance / caused by another factor / light exposure;</p> | <p>if the calculation is omitted from (b)(i) both marks are still available</p> <p>allow ecf from (b)(i)</p> <p>ignore null hypothesis unless explained</p> | [2] |
| (c) (i) | <p>ref. to counting the number of eye piece graticule units;</p> <p>ref. to idea of finding the value of an eyepiece unit with a stage micrometer</p>   | <p>allow descriptions / ref. to a standard graticule unit value</p>   | [2] |

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| <b>(ii)</b>        | <p>Marks are for conclusions about the adaptations shown by the plants. Do not allow marks for answers that restate the data in table 1.1</p> <p>3 of:</p> <p><b>EITHER</b> for shade leaves:</p> <ol style="list-style-type: none"> <li>1. thinner cuticle increases light penetration;</li> <li>2. thinner leaf / shorter palisade cells increases light penetration (to inner parts of leaf);</li> <li>3. spongy mesophyll has more chloroplasts to increase light absorption;</li> <li>4. cells less densely packed / larger air spaces for better gas diffusion;</li> <li>5. larger surface area to absorb limited light / enables more photosynthesis with less light availability;</li> </ol> <p><b>OR</b> for sun / exposed leaves:</p> <ol style="list-style-type: none"> <li>1. thicker cuticle limits water loss;</li> <li>2. (large / long palisade cells) contain more chloroplasts to absorb maximum light;</li> <li>3. fewer chloroplasts in spongy mesophyll as little light penetrates / palisade is light saturated;</li> <li>4. densely packed cells / smaller air spaces reduce water loss;</li> <li>5. smaller surface area reduces water loss;</li> </ol> | <p>Look for the understanding that shade leaves have adaptations that maximise photosynthesis and sun leaves have adaptations to minimise water loss.</p> <p>allow mix and match for sun and shade leaves but take care not to give the same mark twice. candidates should make it clear which type of leaf they are referring to.</p> <p>ignore anything related to growth<br/>ignore any references to internodes<br/>ignore any references to stomata</p> | [3] |
| <b>[Total: 20]</b> |   |  |     |

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| <b>2 (a) (i)</b> | 2 of:<br>length of organism;<br>time for adjustment (to temperature);<br>time of measurement;  | do not allow size   | [2] |
| <b>(ii)</b>      | 1 of:<br>activity / age / sex / mass of organism;<br>source / type / pH / volume of water;<br>oxygen supply;   | do not allow microscope lamp / light  | [1] |
| <b>(iii)</b>     | 1 of:<br>counting high rates is error prone;<br>changes in temperature;<br>activity / stress affect heart rate;  | allow oxygen content if not in <b>(ii)</b><br>e.g. light from microscope / cooling  | [1] |
| <b>(iv)</b>      | idea of sufficient measurements for reliability / to remove anomalous results;   | do not allow accurate<br>ignore reduce error / fair test  | [1] |
| <b>(b) (i)</b>   | allows for different starting points between individuals / can see the changes more clearly;   | looking for the idea that 'it is easier to see what is happening / make comparisons'  | [1] |
| <b>(ii)</b>      | rate at 30 °C – rate at 25 °C / difference in rate at 30 °C and 25 °C<br>rate at 25 °C<br>x 100;   | allow correct use of any figures from the table<br>e.g. $\frac{165 - 132}{132} \times 100$                                      | [1] |
| <b>(iii)</b>     | decrease (by at least 50%) / falls to a very low value / may stop;   |   | [1] |
| <b>(c)</b>       | <i>support:</i><br>5 °C – 15 °C / 15 °C – 25 °C / 10 °C – 20 °C (as rate approx. doubles with 10 °C increase);<br><i>does not support:</i><br>20 °C – 30 °C increases but does not double / 25 °C – 35 °C decreases / above 30 °C rapidly decreases with temperature increase; | allow 'below 30 °C' / 'up to 25 °C'<br><br>allow <u>above</u> 30 °C – <u>35 °C</u> rapidly decreases with temperature increase; | [2] |
|                  |  | <b>[Total: 10]</b>  |     |